#### **ASTA ANALYTICAL METHODS**

Method 15.1

## Volatile Oil in Mustard Seed and Flour (Alternate Method)

Purpose: To determine the volatile oil present in mustard seed and flour. (Gas Chromatographic Method)

## A. Apparatus:

- 1. Gas chromatograph.
  - a. Column, 10' X 1/8" stainless steel packed with 10% Carbowax 20 M and 5% DEGS on A.W. Chromosorb W 60/80 mesh. (Note 1)
  - b. Temperature-Column 150°C; Detector 250°C; Injector 250°C.
  - c. Carrier Gas-Helium at 50 ml/min.
- 2. Recorder or integrator.
- 3. Graduated centrifuge tube 50 mL, with glass stopper.
- 4. High speed coffee grinder.
- 5. Shaker table.
- 6. Vial 10 mL with cap.
- 7. Glass wool, Pasteur pipet.

#### **B.** Reagents:

- 1. Carbon disulfide (CS<sub>2</sub>), ACS grade. (Note 2)
- 2. Allyl isothiocyanate (AIC), ACS grade.

#### C. Preparation of Sample:

- 1. Mustard Flour:
  - a. Weight 1.0 g flour into a centrifuge tube.
  - b. Add 25 mL distilled water.
  - c. Add 10 mL CS<sub>2</sub>.
  - d. Stopper with greased stopper and mix one hour on shaker table.
  - e. Go to D.

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#### 2. Mustard Seed:

- a. Grind seed to pass 30 mesh sieve.
- b. Weigh 1.0 g ground seed into a centrifuge tube.
- c. Add 25 mL boiling distilled water, a little anti-foam and boil 5 more minutes, then cool to room temperature. Add 0.2 g yellow flour.
- d. Add 10 mL CS<sub>2</sub>.
- e. Stopper with greased stopper and mix two hours on shaker table.
- f. Go to D.

#### D. Procedure:

Prepared Samples from C1 or C2:

- 1. Centrifuge sample at 3,000 rpm for five minutes.
- 2. Filter the CS<sub>2</sub> layer through a Pasteur pipette plugged with a small amount of glass wool at the end.
- 3. Use a clean Pasteur pipette to transfer the CS<sub>2</sub> layer of each sample to a 10 mL vial. Allow the CS<sub>2</sub> layer to pass through the wool plug. (Note 3)
- 4. Inject 5  $\mu$ l of CS<sub>2</sub> filtrate or adequate volume.
- 5. When sample has eluted, inject 5  $\mu$ l of standard (Std.). (Note 4)
- 6. Keep vial tightly capped when not in use.

#### E. Calculation:

Relative area = 
$$\frac{\text{Sample area}}{\text{Std. Area}}$$
% Volatile Oil = 
$$\frac{\text{Rel. area X 10 mL extract X } \mu \text{L AIC/mL std.}}{\text{Sample wt. X 1000}} \times 100$$

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= Rel. area XµL AIC/mL std.

Sample wt.

#### F. Statistics:

**TBD** 

#### G. Notes:

- Another column that can be used for this analysis: Column 6' X ¼" glass packed with 20% Carbowax 20 M on A.W. Chromosorb W 60/80 mesh.
- 2. Do all work with carbon disulfide in a well ventilated area of fume hood.
- 3. Filtering the sample prevents mustard from clogging the syringe.
- 4. Standard should be made fresh daily (AIC) approximately the same concentration as the extract--usually 0.2-0.1  $\mu$ L/mL.

### H. References:

AOAC Official Methods of Analysis (1995) 43.1.15 (970.55). JAOAC 4,525 (1921); 53,1 (1970).